

# Enantioselective Total Synthesis of Fluvirucin B<sub>1</sub>

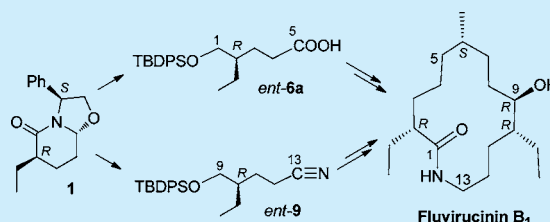
Guillaume Guignard,<sup>†</sup> Núria Llor,<sup>†</sup> Elies Molins,<sup>‡</sup> Joan Bosch,<sup>\*,†</sup> and Mercedes Amat<sup>\*,†</sup>

<sup>†</sup>Laboratory of Organic Chemistry, Faculty of Pharmacy, and Institute of Biomedicine (IBUB), University of Barcelona, 08028 Barcelona, Spain

<sup>‡</sup>Institut de Ciència de Materials (CSIC), Campus UAB, 08193 Cerdanyola, Spain

## S Supporting Information

**ABSTRACT:** A convergent synthesis of fluvirucin B<sub>1</sub> from acid *ent*-6a and nitrile *ent*-9, involving an organocopper coupling, a stereoselective allylation, a ring-closing metathesis reaction, and a stereoselective hydrogenation as the key steps, is reported. The starting building blocks have been prepared in a straightforward manner from a common phenylglycinol-derived lactam **1**. An unprecedented regioselective oxidation of phenylglycinol-derived secondary amines **5** to carboxylic acids **6** has been developed.



Fluvirucins are 14-membered macrocyclic lactams isolated<sup>1–3</sup> from the fermentation broth of actinomycete strains. They are glycosides characterized by the presence of an aminosugar moiety (L-mycosamine, its 4-epimer, or an *N*-substituted derivative) attached at the C-3 or C-9 positions of the core lactam nucleus through a hydroxy group. They also incorporate a methyl or ethyl substituent at the C-2 (1*S*-hydroxyethyl in fluvirucin A<sub>2</sub>), C-6 (absent in some members), and C-10 positions (Figure 1). Fluvirucins possess important and varied

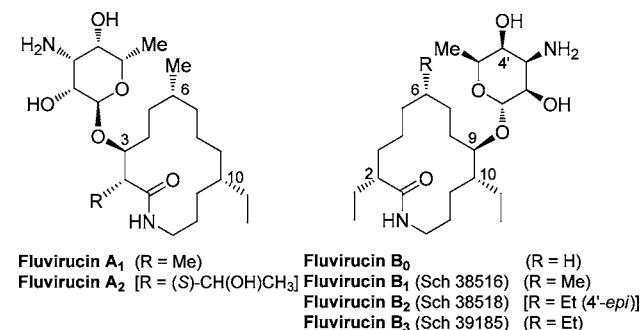


Figure 1. Representative fluvirucins.

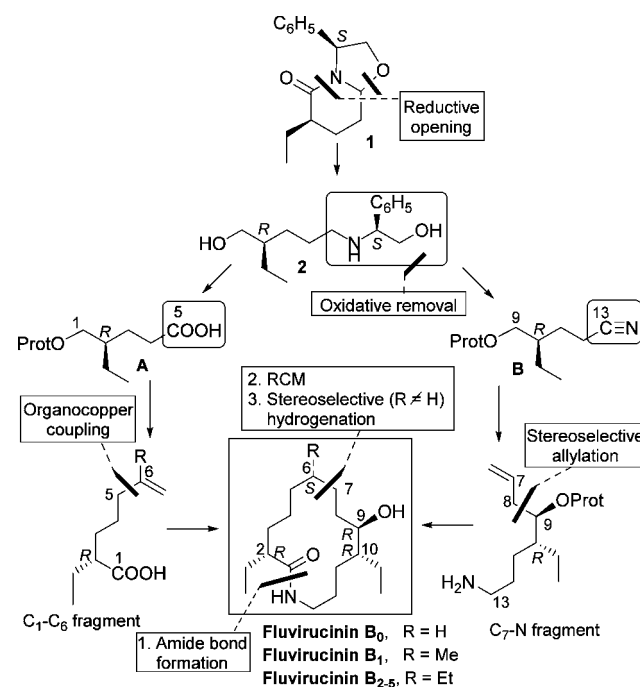
biological activities, such as antifungal,<sup>1</sup> antibiotic,<sup>2</sup> antiviral,<sup>2</sup> and anthelmintic.<sup>3</sup> In particular, fluvirucin B<sub>1</sub> (Sch 38516) exhibits potent antifungal<sup>1a,c</sup> and anti-influenza virus<sup>2a</sup> activities,<sup>4</sup> the latter partially retained in the corresponding aglycon fluvirucin B<sub>1</sub>.<sup>2b</sup>

Although only one total synthesis of a fluvirucin has been reported,<sup>5</sup> the synthesis of the macrocyclic aglycons of fluvirucins, known as fluvirucinins, has received more attention.<sup>6–10</sup> A key point in the synthesis of fluvirucinins is the stereocontrolled assembly of the stereocenters on the macrocyclic ring. Taking into account that all fluvirucinins B possess the same substitution and stereochemical patterns at the C-2 (*R*-Et), C-9 (*S*-OH), and C-10 (*R*-Et) positions, differing only in the C-6 substituent (none in fluvirucin B<sub>0</sub>, *S*-Me in B<sub>1</sub>, *S*-Et

in B<sub>2–5</sub>), we envisaged a unified synthetic strategy to these macrolactams in which the C-2 and C-10 ethyl substituents would come from a common enantiopure amino diol **2**, easily accessible by reductive opening of oxazolopiperidone lactam **1**.<sup>11</sup> Scheme 1 outlines our synthetic plan.

Amino diol **2** would be converted to a 5-hydroxypentanoic acid derivative **A** by oxidative removal of the phenylglycinol moiety and then to the C<sub>1</sub>–C<sub>6</sub> fragment of fluvirucinins B by

## Scheme 1. Unified Synthetic Strategy to Fluvirucinins B



Received: February 23, 2016

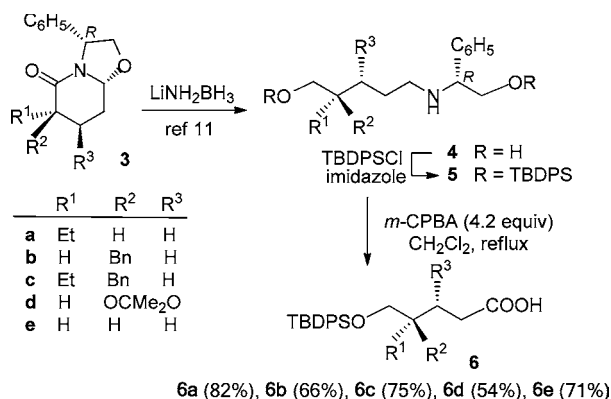
Published: April 5, 2016

copper-catalyzed coupling of the corresponding iodide with an appropriately substituted ( $R = H, Me$  or  $Et$ ) alkenyl Grignard reagent. In turn, the secondary amino group of amino diol **2** would be oxidized to a cyano group, and the resulting 5-hydroxypentanenitrile **B** would be converted to the  $C_7$ -N fragment of fluvirucinins **B** after the incorporation of the  $C_9$  stereogenic center by a stereoselective allylation of an aldehyde. Linkage of the two fragments by an amidation reaction, followed by a ring-closing metathesis and stereoselective hydrogenation of the resultant alkene, would complete the synthesis of the target fluvirucinins **B**. The success of our synthetic plan would rely on the development of efficient procedures for the oxidative removal of the phenylglycinol moiety present in amino diol **2** to afford 5-hydroxypentanoic acid and 5-hydroxypentanenitrile derivatives.

The conversion of a secondary amine to a carboxylic acid is a challenging, unprecedented transformation. Taking into account that primary amines are oxidized to nitro derivatives by treatment with *m*-chloroperbenzoic acid,<sup>12</sup> we decided to study this oxidation using a set of phenylglycinol-derived secondary amines structurally related to **2**.

To our delight, treatment of the *O*-protected amino diols **5a–d** with an excess of *m*-CPBA (4.2 equiv) in refluxing  $CH_2Cl_2$  directly afforded the corresponding carboxylic acids **6a–d**, bearing a variety of substituents (ethyl, benzyl, isopropylidenedioxy) in good yields (Scheme 2). Considering

**Scheme 2. Oxidative Removal of the Chiral Inductor: Access to Enantiopure *O*-Protected 5-Hydroxypentanoic Acids**



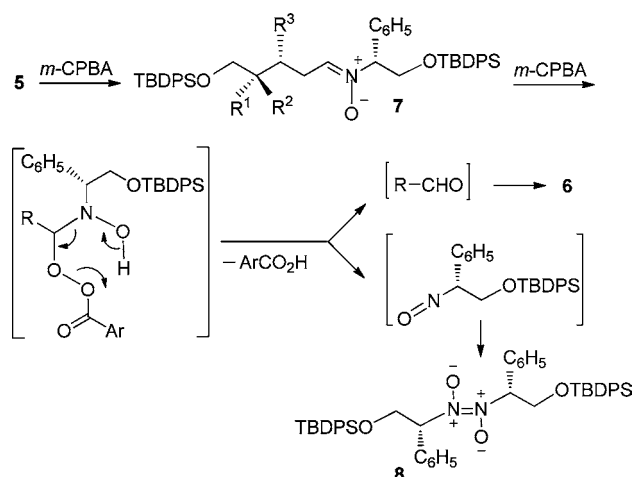
that amino diols **4** are available with virtually any type of substitution pattern,<sup>11</sup> the above oxidative procedure opens a general synthetic entry to enantiopure 5-hydroxypentanoic acid derivatives.

The formation of carboxylic acids **6** can be accounted for by considering the generation of the nonconjugated nitrones **7**<sup>13</sup> and their *m*-CPBA-promoted oxidative cleavage<sup>14</sup> with subsequent oxidation of the resulting aldehyde. The oxidative cleavage also produces a nitroso derivative, which was isolated as the corresponding nitroso dimer **8** (Scheme 3).

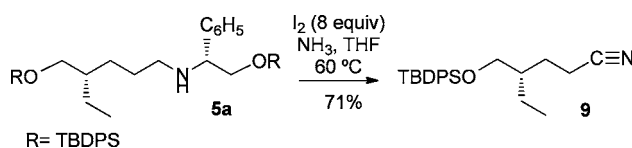
In support of this mechanism, nitron **7e**, prepared by  $Na_2WO_4$ /hydrogen peroxide–urea oxidation<sup>13b</sup> of the simple secondary amine **5e**, was converted to hydroxypentanoic acid derivative **6e** (45% from **5e**) and dimer **8** by treatment with *m*-CPBA (2.5 equiv).

On the other hand, the generation of a 5-hydroxypentanenitrile by oxidative cleavage of the phenylglycinol moiety of the starting *O*-protected amino diol was successfully accomplished in a single step using molecular iodine in aqueous ammonia.<sup>15</sup>

**Scheme 3. Proposed Mechanism for the *m*-CPBA-Promoted Oxidation of Secondary Amines **5****



**Scheme 4. Oxidation of Secondary Amine **5a** to Nitrile **9****



In this way, secondary amine **5a** was converted to nitrile **9** in 71% yield (Scheme 4).

This transformation involves the initial generation of an imine and its reaction with ammonia to form an aminal, which decomposes to a primary amine and an imine. Subsequent oxidation and hydrolytic steps lead to the nitrile and (*tert*-butyldiphenylsilyloxy)methyl phenyl ketone, regardless of the regioselectivity of the initial oxidation.

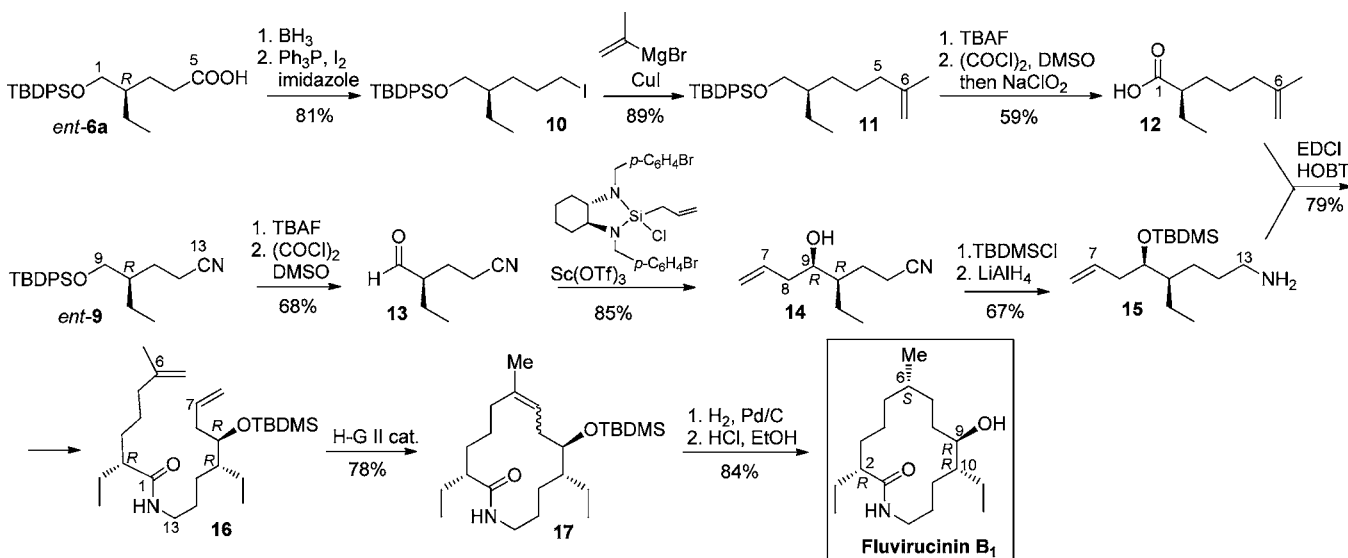
Having developed straightforward procedures for the conversion of secondary amines **5** to functionalized carboxylic acids **6** and nitrile **9**, to evaluate the feasibility of the unified strategy outlined in Scheme 1, we undertook the synthesis of fluvirucinins **B**. To achieve the required 2*R* and 10*R* configuration characteristic of fluvirucinins **B**, we started from the (*S*)-phenylglycinol-derived secondary amine *ent*-**4a** (= **2**), which was converted, as in the above (*R*)-phenylglycinol series, to hydroxy acid *ent*-**6a** (**A**; Prot = TBDPS) and hydroxy nitrile *ent*-**9** (**B**; Prot = TBDPS).

Scheme 5 outlines the synthesis of fluvirucinins **B**. The  $C_1$ – $C_6$  fragment (compound **12**) was prepared from carboxylic acid *ent*-**6a**, which was converted, via an alcohol, to iodide **10**.

A subsequent cross-coupling with 2-propenylmagnesium bromide in the presence of a catalytic amount of  $CuI$ <sup>16</sup> (bond formed  $C_5$ – $C_6$ ) provided the protected alcohol **11**, which was desilylated and oxidized to carboxylic acid **12** (23% overall yield from **1**).

On the other hand, after the protected hydroxy nitrile *ent*-**9** was converted to aldehyde **13**, a stereoselective allylation using the (*S,S*)-Leighton reagent<sup>17</sup> installed the  $C_9$  stereogenic center to give homoallylic *syn* alcohol **14**<sup>18</sup> (bond formed  $C_8$ – $C_9$ ). Protection of the hydroxy group of **14**, followed by reduction of the cyano group, afforded amine **15** (the  $C_7$ –N fragment of fluvirucinins **B**) in 21% overall yield from **1**.

Coupling of the two fragments, carboxylic acid **12** and amine **15**, took place in excellent yield to give amide **16**. A subsequent ring-closing metathesis reaction (bond formed  $C_6$ – $C_7$ ),

Scheme 5. Total Synthesis of Fluvirucin B<sub>1</sub><sup>a</sup>

<sup>a</sup>The carbon numbering of the intermediates corresponds to that of fluvirucin B<sub>1</sub>.

followed by stereoselective catalytic hydrogenation of the resulting 1.2:1 mixture of trisubstituted olefins 17, generated the C-6 stereocenter of the macrocycle,<sup>19</sup> leading to the O-protected fluvirucin derivative 18. The NMR data of our silyl derivative 18 matched those reported in the literature,<sup>5b,9b</sup> and its mp and absolute rotation were consistent with those previously reported.<sup>9b</sup> Additionally, the absolute configuration of 18 was unambiguously established by X-ray crystallographic analysis<sup>20</sup> (Figure 2). A final removal of the silyl protecting group

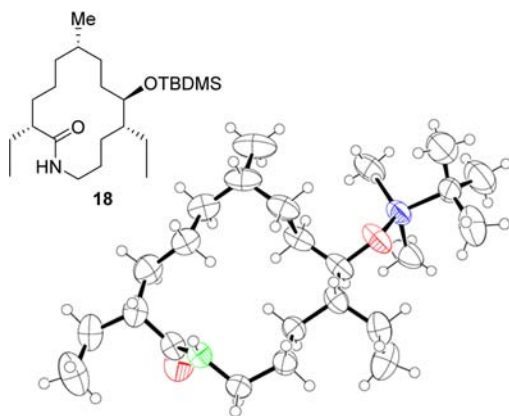


Figure 2. X-ray crystal structure of the fluvirucin B<sub>1</sub> precursor 18.

completed the synthesis of fluvirucin B<sub>1</sub>, whose NMR data and [α] value are reported for the first time (see the Supporting Information).

The convergent enantioselective synthesis of fluvirucin B<sub>1</sub> herein reported consists of 12 linear synthetic steps from phenylglycinol-derived lactam 1<sup>21</sup> in the longest linear sequence. The overall yield was 11%, which compares advantageously with previous syntheses<sup>9</sup> of this aglycon. The synthesis also features an unprecedented oxidation of phenylglycinol-derived secondary amines 5 to diversely substituted enantiopure 5-hydroxypentanoic acid derivatives 6. By using an appropriate alkenyl Grignard reagent in the assembly of the C<sub>1</sub>–C<sub>6</sub> fragment, the strategy we have developed could be applied to the synthesis of fluvirucins B<sub>0</sub> and B<sub>2–5</sub>.

## ■ ASSOCIATED CONTENT

### § Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.6b00513.

Experimental procedures, product characterizations, and <sup>1</sup>H and <sup>13</sup>C NMR spectra (PDF)

Crystallographic data for compound 18 (CIF)

## ■ AUTHOR INFORMATION

### Corresponding Authors

\*E-mail: joanbosch@ub.edu.

\*E-mail: amat@ub.edu.

### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

Financial support from the Spanish Ministry of Economy and Competitiveness/FEDER (Projects CTQ2012-35250 and CTQ2015-65384-R) and the Generalitat de Catalunya (Grant No. 2014-SGR-155) is gratefully acknowledged. We also acknowledge the networking contribution from the COST Action CM1407.

## ■ REFERENCES

- (a) Hegde, V. R.; Patel, M. G.; Gullo, V. P.; Ganguly, A. K.; Sarre, O.; Puar, M. S.; McPhail, A. T. *J. Am. Chem. Soc.* **1990**, *112*, 6403–6405. (b) Hegde, V. R.; Patel, M. G.; Gullo, V. P.; Puar, M. S. *J. Chem. Soc., Chem. Commun.* **1991**, 810–812. (c) Hegde, V.; Patel, M.; Horan, A.; Gullo, V.; Marquez, J.; Gunnarsson, I.; Gentile, F.; Loebenberg, D.; King, A.; Puar, M.; Pramanik, B. *J. Antibiot.* **1992**, *45*, 624–632. (d) Cooper, R.; Truumees, I.; Yarborough, R.; Loebenberg, D.; Marquez, J.; Horan, A.; Patel, M.; Gullo, V.; Puar, M.; Pramanik, B. *J. Antibiot.* **1992**, *45*, 633–638.
- (a) Naruse, N.; Tenmyo, O.; Kawano, K.; Tomita, K.; Ohgusa, N.; Miyaki, T.; Konishi, M.; Oki, T. *J. Antibiot.* **1991**, *44*, 733–740. (b) Naruse, N.; Tsuno, T.; Sawada, Y.; Konishi, M.; Oki, T. *J. Antibiot.* **1991**, *44*, 741–755. (c) Naruse, N.; Konishi, M.; Oki, T.; Inouye, Y.; Kakisawa, H. *J. Antibiot.* **1991**, *44*, 756–761.

- (3) (a) Ayers, S.; Zink, D. L.; Mohn, K.; Powell, J. S.; Brown, C. M.; Murphy, T.; Grund, A.; Genilloud, O.; Salazar, O.; Thompson, D.; Singh, S. B. *J. Nat. Prod.* **2007**, *70*, 1371–1373. (b) Ayers, S.; Zink, D. L.; Powell, J. S.; Brown, C. M.; Grund, A.; Genilloud, O.; Salazar, O.; Thompson, D.; Singh, S. B. *J. Antibiot.* **2008**, *61*, 59–62.
- (4) (a) For the biosynthesis of the aglycons of fluvirucins B<sub>1</sub>, B<sub>2</sub>, and B<sub>3</sub>, see: Puar, M. S.; Gullo, V.; Gunnarsson, I.; Hegde, V.; Patel, M.; Schwartz, J. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 575–578. (b) For the identification and characterization of the fluvirucin B<sub>1</sub> polyketide synthase, see: Lin, T.-Y.; Borketey, L. S.; Prasad, G.; Waters, S. A.; Schnarr, N. A. *ACS Synth. Biol.* **2013**, *2*, 635–642.
- (5) Fluvirucin B<sub>1</sub>: (a) Xu, Z.; Johannes, C. W.; Salman, S. S.; Hoveyda, A. H. *J. Am. Chem. Soc.* **1996**, *118*, 10926–10927. (b) Xu, Z.; Johannes, C. W.; Hour, A. F.; La, D. S.; Cogan, D. A.; Hofilena, G. E.; Hoveyda, A. H. *J. Am. Chem. Soc.* **1997**, *119*, 10302–10316.
- (6) Fluvirucin A<sub>1</sub>: (a) Suh, Y.-G.; Kim, S.-A.; Jung, J.-K.; Shin, D.-Y.; Min, K.-H.; Koo, B.-A.; Kim, H.-S. *Angew. Chem., Int. Ed.* **1999**, *38*, 3545–3547. (b) Liang, B.; Negishi, E. *Org. Lett.* **2008**, *10*, 193–195. (c) Son, S.; Fu, G. C. *J. Am. Chem. Soc.* **2008**, *130*, 2756–2757 (formal). (d) Radha Krishna, P.; Anitha, K. *Tetrahedron Lett.* **2011**, *52*, 4546–4549. (e) Suh, Y.-G.; Lee, Y.-S.; Kim, S.-H.; Jung, J.-K.; Yun, H.; Jang, J.; Kim, N.-J.; Jung, J.-W. *Org. Biomol. Chem.* **2012**, *10*, 561–568.
- (7) Fluvirucin A<sub>2</sub>: Lee, Y.-S.; Jung, J.-W.; Kim, S.-H.; Jung, J.-K.; Paek, S.-M.; Kim, N.-J.; Chang, D.-J.; Lee, J.; Suh, Y. G. *Org. Lett.* **2010**, *12*, 2040–2043.
- (8) Fluvirucin B<sub>0</sub>: Baltrusch, A. W.; Bracher, F. *Synlett* **2002**, 1724–1726.
- (9) Fluvirucin B<sub>1</sub>: (a) Hour, A. F.; Xu, Z.; Cogan, D. A.; Hoveyda, A. H. *J. Am. Chem. Soc.* **1995**, *117*, 2943–2944. (b) Trost, B. M.; Ceschi, M. A.; König, B. *Angew. Chem., Int. Ed. Engl.* **1997**, *36*, 1486–1489. (c) Martín, M.; Mas, G.; Urpí, F.; Vilarrasa, J. *Angew. Chem., Int. Ed.* **1999**, *38*, 3086–3089. See also ref 5b.
- (10) Fluvirucin B<sub>2-5</sub>: Llàcer, E.; Urpí, F.; Vilarrasa, J. *Org. Lett.* **2009**, *11*, 3198–3201.
- (11) Guignard, G.; Llor, N.; Urbina, A.; Bosch, J.; Amat, M. *Eur. J. Org. Chem.* **2016**, 2016, 693–703.
- (12) Gilbert, K. E.; Borden, W. T. *J. Org. Chem.* **1979**, *44*, 659–661.
- (13) For the preparation of nitrones by oxidation of secondary amines, see: (a) Murahashi, S.-I.; Mitsui, H.; Shiota, T.; Tsuda, T.; Watanabe, S. *J. Org. Chem.* **1990**, *55*, 1736–1744. (b) Marcantoni, E.; Petrini, M.; Polimanti, O. *Tetrahedron Lett.* **1995**, *36*, 3561–3562. (c) Colonna, S.; Pironti, V.; Carrea, G.; Pasta, P.; Zambianchi, F. *Tetrahedron* **2004**, *60*, 569–575. (d) Gella, C.; Ferrer, E.; Alibés, R.; Busqué, F.; de March, P.; Figueredo, M.; Font, J. *J. Org. Chem.* **2009**, *74*, 6365–6367.
- (14) For peracid-promoted oxidative ring-opening of cyclic nitrones, see: Bapat, J. B.; Durie, A. *Aust. J. Chem.* **1984**, *37*, 211–219.
- (15) (a) Iida, S.; Togo, J. *Tetrahedron* **2007**, *63*, 8274–8281. See also: (b) Veisi, H. *Synthesis* **2010**, 2631–2635. (c) Zhu, C.; Sun, C.; Wei, Y. *Synthesis* **2010**, 4235–4241.
- (16) For copper-catalyzed couplings of alkenyl Grignard reagents with primary alkyl iodides, see: (a) Derguini-Boumechal, F.; Linstumelle, G. *Tetrahedron Lett.* **1976**, *17*, 3225–3226. For more recent examples, see: (b) Takahashi, M.; Dodo, K.; Hashimoto, Y.; Shirai, R. *Tetrahedron Lett.* **2000**, *41*, 2111–2114. (c) Kochi, T.; Ellman, J. A. *J. Am. Chem. Soc.* **2004**, *126*, 15652–15653. (d) Terayama, N.; Yasui, E.; Mizukami, M.; Miyashita, M.; Nagumo, S. *Org. Lett.* **2014**, *16*, 2794–2797. For the use of iron catalysts, see: (e) Guérinot, A.; Reymond, S.; Cossy, J. *Angew. Chem., Int. Ed.* **2007**, *46*, 6521–6524. (f) Cahiez, G.; Duplais, C.; Moyeux, A. *Org. Lett.* **2007**, *9*, 3253–3254.
- (17) (a) Kubota, K.; Leighton, J. L. *Angew. Chem., Int. Ed.* **2003**, *42*, 946–948. For synthetic applications, see: (b) Vintonyak, V. V.; Maier, M. E. *Org. Lett.* **2008**, *10*, 1239–1242. (c) Harsh, P.; O'Doherty, G. A. *Tetrahedron* **2009**, *65*, 5051–5055 and references cited therein. (d) For the use of Sc(OTf)<sub>3</sub> as a catalyst in enantioselective Leighton allylations, see: Kim, H.; Ho, S.; Leighton, J. L. *J. Am. Chem. Soc.* **2011**, *133*, 6517–6520. See also ref 10.
- (18) Minor amounts (dr = 9:1) of the *anti* adduct were detected by NMR. Purification of amine **15** afforded a single diastereoisomer.
- (19) A similar remote macrocyclic stereocontrol in the synthesis of fluvirucinins was first observed by Hoveyda<sup>5</sup> in the hydrogenation of related macrocyclic olefins bearing a trisubstituted C<sub>5</sub>–C<sub>6</sub> (instead of C<sub>6</sub>–C<sub>7</sub>) double bond.
- (20) CCDC 1440667 contains the supplementary crystallographic data for compound **18**. This data can be obtained free of charge from the Cambridge Crystallographic Data Centre via [www.ccdc.cam.ac.uk/data\\_request/cif](http://www.ccdc.cam.ac.uk/data_request/cif).
- (21) For recent enantioselective total syntheses of complex alkaloids using amino alcohol-derived lactams as starting materials, see: (a) Amat, M.; Ramos, C.; Pérez, M.; Molins, E.; Florindo, P.; Santos, M. M. M.; Bosch, J. *Chem. Commun.* **2013**, 49, 1954–1956. (b) Ballette, R.; Pérez, M.; Proto, S.; Amat, M.; Bosch, J. *Angew. Chem., Int. Ed.* **2014**, *53*, 6202–6205. (c) Amat, M.; Guignard, G.; Llor, N.; Bosch, J. *J. Org. Chem.* **2014**, *79*, 2792–2802. (d) Amat, M.; Pinto, A.; Griera, R.; Bosch, J. *Chem. - Eur. J.* **2015**, *21*, 12804–12808. For reviews, see: (e) Amat, M.; Pérez, M.; Bosch, J. *Synlett* **2011**, 2011, 143–160. (f) Amat, M.; Llor, N.; Griera, R.; Pérez, M.; Bosch, J. *Nat. Prod. Commun.* **2011**, *6*, 515–526.